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Association of APOE-Independent Alzheimer Disease Polygenic Risk Score With Brain Amyloid Deposition in Asymptomatic Older Adults

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Author(s):

Laura Xicota, PhD¹; Beata Gyorgy, Msc¹; Benjamin Grenier-Boley, Msc.²; Alexandre Lecoeur, Msc.¹; Gaëlle Fontaine, Msc.¹; Fabrice Danjou, PhD¹; Jorge Samper Gonzalez, PhD¹,³; Olivier Colliot, PhD¹,³; Philippe Amouyel, MD PhD²; Garance Martin, Msc⁴; Marcel Levy, MD⁵; Nicolas Villain, MD PhD¹,6; Marie-Odile Habert, MD PhD⁴,5,7,8; Bruno Dubois, MD PhD¹,6,5; Jean-Charles Lambert, PhD²; Marie-Claude Potier, PharmD PhD¹ on behalf of the INSIGHT pre-AD study group and for the Alzheimer's Disease Neuroimaging Initiative*

Corresponding Author:

Marie-Claude Potier, marie-claude.potier@upmc.fr

Affiliation Information for All Authors: 1. ICM Paris Brain Institute, CNRS UMR7225, INSERM U1127, Sorbonne University, Hôpital de la Pitié-Salpêtrière, 47 Bd de l'Hôpital, 75013 Paris, France; 2. Univ. Lille, Inserm, CHU Lille, Institut Pasteur de Lille, U1167-RID-AGE – Facteurs de risque et déterminants moléculaires des maladies liées au vieillissement, F-59000 Lille, France; 3. Inria, Aramis-project team, Paris, France; 4. Centre d'Acquisition et Traitement des Images (CATI platform), www.cati-neuroimaging.com; 5. Centre des Maladies Cognitives et Comportementales, IM2A, AP-HP, Sorbonne Université, Hôpital de la Salpêtrière, Paris, France; 6. AP-HP Sorbonne Université, Hôpital Pitié-Salpêtrière, Department of Neurology, Paris, France; 7. Sorbonne Université, CNRS, INSERM, Laboratoire d'Imagerie Biomédicale, LIB, F-75006, Paris, France; 8. AP-HP, Hôpital Pitié-

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Salpêtrière, Médecine Nucléaire, F-75013, Paris, France

Equal Author Contribution:

Contributions:

Laura Xicota: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data; Study concept or design; Analysis or interretation of data

Beata Gyorgy: Drafting/revision of the manuscript for content, including medical writing for content; Analysis or interretation of data

Benjamin Grenier-Boley: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data; Analysis or interretation of data Alexandre Lecoeur: Drafting/revision of the manuscript for content, including medical writing for content; Analysis or interretation of data

Gaëlle Fontaine: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data

Fabrice Danjou: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data; Analysis or interpretation of data

Jorge Samper Gonzalez: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data

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Olivier Colliot: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data

Philippe Amouyel: Drafting/revision of the manuscript for content, including medical writing for content

Garance Martin: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data

Marcel Levy: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data

Nicolas Villain: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data; Analysis or interretation of data

Marie-Odile Habert: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data; Study concept or design; Analysis or interretation of data

Bruno Dubois: Drafting/revision of the manuscript for content, including medical writing for content; Study concept or design

Jean-Charles Lambert: Drafting/revision of the manuscript for content, including medical writing for content; Analysis or interretation of data

Marie-Claude Potier: Drafting/revision of the manuscript for content, including medical writing for content; Major role in the acquisition of data; Study concept or design; Analysis or interretation of data

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Abstract

Background and objectives: Brain amyloid deposition, a major risk factor for Alzheimer's disease (AD), is currently estimated by measuring cerebrospinal fluid or plasma amyloid peptide levels, or by positron-emission tomography imaging. Assessing genetic risks relating to amyloid

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deposition before any accumulation has occurred would allow for earlier intervention in persons at increased risk for developing AD. Previous work linking amyloid burden and genetic risk relied almost exclusively on *APOE*, a major AD genetic risk factor. Here, we ask whether a polygenic risk score (PRS) that incorporates an optimized list of common variants linked to AD and excludes *APOE* is associated with brain amyloid load in cognitively unimpaired elderly adults.

Methods: We included 291 elderly asymptomatic participants from the INveStIGation of AlzHeimer's PredicTors (INSIGHT-preAD) cohort who underwent amyloid imaging, including 83 amyloid-positive (+) participants. We used an Alzheimer's (A) PRS composed of 33 AD risk variants excluding *APOE*, and selected the 17 variants that showed the strongest association with amyloid positivity to define an optimized (oA) PRS. Participants from the Alzheimer's Disease Neuroimaging Initiative (ADNI) study [228 participants, 90 amyloid (+)] were tested as a validation cohort. Finally, 2,300 AD patients and 6,994 controls from the European Alzheimer's Disease Initiative (EADI) were evaluated.

Results: A-PRS was not significantly associated with amyloid burden in the INSIGHT or ADNI cohorts with or without correction for *APOE* genotype. However, oA-PRS was significantly associated with amyloid status independently of *APOE* adjustment (INSIGHT OR: 5.26 [1.71-16.88]; ADNI OR: 3.38 [1.02-11.63]). Interestingly, oA-PRS accurately discriminated amyloid (+) and (-) *APOE* ε4 carriers (INSIGHT OR: 181.6 [7.53-10,674.6]; ADNI OR: 44.94 [3.03-1,277]). A-PRS and oA-PRS showed a significant association with disease status in the EADI cohort (OR: 1.68 [1.53-1.85] and 2.06 [1.73-2.45] respectively). Genes assigned to oA-PRS variants were enriched in ontologies related to Aβ metabolism and deposition.

Discussion: PRSs relying on AD genetic risk factors excluding *APOE* may improve risk prediction for brain amyloid, allowing stratification of cognitively unimpaired individuals at risk of AD independent of their *APOE* status.



1. Background

Alzheimer's disease (AD) is the most common cause of dementia and a major public health concern, with >130 million cases worldwide anticipated by 2050. AD is a complex disease with autosomal dominant transmission in rare early-onset familial AD¹ and a non-Mendelian inheritance pattern in late-onset sporadic AD (sAD) that may explain 60-80% of the attributable risk². The first identified genetic variant associated with AD was the apolipoprotein E (*APOE*) ε4 allele³. Heterozygous carriers have three-fold higher AD risk, while homozygous individuals have 15-fold higher AD risk⁴. The AD risk for homozygous individuals is estimated to be 30% at age 75 and over 50% by age 85⁵. Since 2009, genome wide association studies (GWAS) have identified more than 40 loci associated with sAD⁴.6.

Notably, the risk of developing AD associated with these GWAS variants is low, and therefore it is of interest to calculate a weighted sum of identified risk variants to establish the cumulative risk of disease or phenotypic trait for a given individual, known as a polygenic risk score (PRS). Such approaches have been used to differentiate AD-related dementia stages⁷⁻¹¹ and to predict age of disease onset¹²⁻¹⁴ and/or clinical progression⁷⁻¹¹. In some cases, the association was dependent on the *APOE* genotype^{7,12}.

Few studies focused on the association of PRS with relevant AD-linked phenotypes in cognitively unimpaired older adults. Mormino *et al.*¹⁵ reported that, in participants without dementia, PRS was associated with cerebral accumulation of A β measured by positron emission tomography (PET). Similarly, a study of middle-aged individuals with a familial history of sAD revealed that specific PRSs that included *APOE* were associated with PET and cerebrospinal fluid (CSF) amyloid load¹⁶. A recent study based on the Alzheimer's Disease Neuroimaging Initiative (ADNI) cohort separated participants with AD-associated dementia, AD-associated

mild cognitive impairment (MCI), and controls into amyloid (+) and amyloid (-) groups based on amyloid PET. Among the groups, a high-content PRS generated from 162,957 single-nucleotide polymorphisms (SNPs) did not predict amyloid status better than the *APOE* genotype alone¹⁰. Nevertheless, when using pathway-specific PRSs, lists related to lipid-protein interactions and cholesterol transport were significantly associated with brain amyloid load, even when excluding $APOE^{10}$. Finally, a recent study using 39 AD genetic variants found that a high PRS and $APOE \, \varepsilon 4$ separately predicted AD dementia in a retrospective cohort¹⁷.

Here, our objective was to test whether we could generate a PRS linked to amyloid status in cognitively unimpaired participants using a list of SNPs previously associated with AD but excluding *APOE*. A PRS optimized for amyloid status could identify at-risk individuals, encouraging them to seek future targeted prevention efforts.

2. Materials and methods

2.1. Discovery cohort: INSIGHT cohort

We used data from the INveStIGation of AlzHeimer's PredicTors in a subjective memory complainer pre Alzheimer's disease (INSIGHT-preAD) cohort comprising cognitively unimpaired volunteers, aged 70 years and older, who consulted at the Pitié-Salpêtrière University Hospital for memory complaints. All participants included had a Mini-Mental State Examination (MMSE) score ≥27¹⁸, a Dementia Rating Score of 0 and normal episodic memory performance (assessed with the Free and Cued Selective Reminding Test). Additional available data for this population include age, sex, weight, body mass index, *APOE* genotype, medical treatments, education, residence location, as well as extensive neuropsychological and neuro-imaging (MRI and FDG-PET) data. Participants underwent an initial ¹⁸F-florbetapir PET scan to assess their

brain amyloid load and were classified as amyloid (+) or amyloid (-). The global amyloid PET standard uptake value ratio (SUVR) was calculated as described previously¹⁸⁻²⁰. To compare amyloid burden in several large cohorts using different radiotracers and analysis methods, a standardized scale of amyloid burden quantification was proposed by Klunk²¹. This scale goes from 0 to 100, using a new unit called a centiloid (CL). SUVR values were transformed to CL values using the center for acquisition and image Processing (CATI) platform²² by applying a three-level method accounting for the radiotracer and the pipeline used to process the PET amyloid data^{21,23}. INSIGHT participants were then divided into amyloid (-) and amyloid (+) groups using a 20-CL threshold, corresponding to an SUVR value of 0.79 and the following conversion equation: CL = (151* SUVR) – 98.9. A cut-off of 20 CL was previously validated in populations with post-mortem findings^{24,25}. The ethics committee of the Pitié-Salpêtrière University Hospital approved the study protocol. All participants provided written informed consent via a form given and explained to them two weeks before enrollment. Neither the participants nor the investigators were aware of any participant's amyloid status.

2.2. Validation cohort: ADNI cohort

Additional data were obtained from the ADNI database²⁶. The ADNI was launched in 2003 as a public-private partnership, led by Principal Investigator Michael W. Weiner, MD. The primary goal of ADNI is to test whether serial magnetic resonance imaging, PET, other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of MCI and early AD.

The ADNI cohort is an independent cohort including controls and participants with MCI or AD²⁶. We selected control participants from the ADNI cohort who underwent an¹⁸F-

florbetapir PET scan, as with the INSIGHT cohort; had an MMSE score ≥27; and were within the same age range as the INSIGHT cohort. SUVR values from the ADNI were transformed to CL using the following formula: CL = (196.9*SUVR) − 196.03. We used the same threshold for amyloid positivity as for the INSIGHT cohort (20 CL).

2.3. AD study: European Alzheimer's Disease Initiative (EADI)

EADI is composed of several case-control studies and one population-based cohort, 3C²⁷. The case-control studies are comprised of AD cases and cognitively normal controls across France. The population-based cohort is from a prospective study on the relationship between vascular factors and dementia carried out in the three French cities: Bordeaux, Montpellier and Dijon. AD status was defined based on 12 years follow-up for Dijon participants, 14-15 years follow-up for Montpellier participants and 17-18 years follow-up for Bordeaux participants. Non-demented subjects from the 3C cohort were included as controls. All AD cases in the case-control studies and in 3C were ascertained by neurologists and the clinical diagnosis of probable AD was established according to the DSM-III-R and NINCDS-ADRDA criteria²⁸.

2.4. Genotyping

INSIGHT participants were genotyped using the Illumina NeuroX2 chip, a semi-custom microarray based on a HumanCore-24+® v1.0 backbone containing 306,670 variants, with an additional 179,467 custom variants relevant for neurological diseases. The design of this chip was reported previously²⁹. Data quality control was performed using Genome Studio 2.0 software (Illumina) and plink v.1.9 beta³⁰. Quality control filtering removed 21,644 SNPs with low GenTrain scores (<0.7) and low genotyping rates (<98%), as well as those deviating from

Hardy-Weinberg equilibrium (HWE test p-value $< 10^{-6}$). Samples were checked for low call rate (<98%), individual relatedness, and ethnic discrepancies. The inbreeding coefficient was considered excessive when Fhat2<-0.8. Sex discrepancies were checked and data updated where possible. Following these criteria, 10 participants were removed from further analyses. Imputation was performed using the Sanger Imputation Service on the Haplotype Reference Consortium dataset (release 1.1)³¹. Low-imputation-quality variants were filtered using a threshold of $r^2<0.3$.

The ADNI cohort was genotyped using different Illumina microarrays; therefore, quality control and imputation were conducted separately using the same procedures. Variants were filtered for GenTrain score <0.7, clusterSeparation score <=0.3, low call-rate (<99%), rare variants (minor allele frequency <5%) and deviation from HWE with p< 10^{-6} . Samples were filtered for missingness (>2%), relatedness, sex discrepancy, and excess heterozygosity. Imputation was performed using the Sanger Imputation Service on the Haplotype Reference Consortium dataset. Low-imputation-quality variants were filtered at a threshold of r^2 <0.3.

The EADI study cohort was genotyped using the Illumina Human 610 Quad BeadChip at the Centre National de Recherche en Génomique Humaine (CNRGH, Evry, France). The genotyping chip was assessed using probe alignment and a remapping and normalization step according to the GRCh37 and GRCh38 assemblies. Sample quality control was performed as previously detailed³². Relatedness and variant quality control were re-computed as previously described³³. Briefly, variants with a minor allele frequency <0.01, missingness >0.05, a p-value from the Hardy-Weinberg Equilibrium test performed in controls <5e⁻⁸ or a p-value of the Fisher's exact test on cases/controls missing calls <1e⁻¹⁰ were excluded. The remaining variants were then assessed by comparing their frequencies against two reference panels (i.e. the

Haplotype Reference Consortium r1.1 (HRC)³¹ excluding 1000 Genomes samples and the Genome Aggregation Database v3 (gnomAD) non-Finnish European samples³⁴). Allele counts were then compared to the EADI counts by performing a chi-square test (χ^2); variants showing a $\chi^2 > 1,500$ in both HRC and gnomAD, or a $\chi^2 > 1,500$ in one reference panel and not present in the other, were excluded. All samples and variants passing quality control were then imputed with the Trans-Omics for Precision Medicine (TOPMed) Freeze 5 reference panel³⁵ on the Michigan Imputation Server³⁶. Low-imputation-quality variants were filtered at a threshold of $r^2 < 0.3$.

2.5. PRS calculation and statistical analysis

To calculate the Alzheimer's PRS (A-PRS), we used a list of previously described SNPs in Bellenguez *et al.* ⁶ that were confirmed to be linked to AD. SNPs were included only if their allelic frequency was higher than 1% in the population (including *TREM2* rs75932628, *PLCG2* rs72824905, *HESX1/IL17RD/APPL1* rs184384746, *CNTAP2* rs114360492, and *TM2D3* rs139709573). All included SNPs are considered to be sentinel SNPs and were used in the calculation of the A-PRS. Exceptions were rs9271058 and rs12881735, which did not pass quality control in our cohort and were substituted by the closest available SNPs after confirming linkage disequilibrium between them, and rs113260531, for which no odds ratio has been published³⁷ (Table 1).

All statistics and PRS calculations were performed on R 4.0.2.³⁸. Polygenic risk scores were calculated as described previously³⁹, using a weighted method with the following formula: $PRS = \frac{\sum_{n=1}^{nSNP} Dose*ln(OR)}{\sum_{n=1}^{nSNP} ln(OR)}$. Dose varied between 0 and 2, with 0 corresponding to no risk allele, 1 to one risk allele, and 2 to two risk alleles. For imputed alleles, the dose was a continuous value between 0 and 2.

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 χ^2 and Kruskal-Wallis analyses were performed to determine differences in population demographics. All correlations were obtained using the Spearman correlation method. *APOE* status was defined according to the $\epsilon 4$ carrier status of the participant, and only participants with $\epsilon 3/\epsilon 4$ and $\epsilon 4/\epsilon 4$ genotypes were included in the E4 carriers, while all remaining subjects were included among the E4 non-carriers. To calculate the association of the PRS and/or *APOE* group with amyloid status, models were fitted for a binomial response adjusted for age, sex, and the first three principal components of each population to account for the internal structure of the INSIGHT cohort and ten principal components to account for the internal structure of ADNI. Only the first three principal components were included for INSIGHT because it is a homogeneous population, whereas the ADNI cohort is multiethnic. Principal components were calculated using the pca function of PLINK (v1.90b3w), and plotted against the 1000G dataset. Non-European outliers were identified and removed from the INSIGHT cohort. These models were subsequently used to obtain the beta value of the PRS using the R reghelper package. We present uncorrected p-values.

2.6. PRS optimization

The optimized Alzheimer's PRS (oA-PRS) was obtained using the INSIGHT discovery cohort and validated in the ADNI validation cohort. We generated n-1 lists of SNPs excluding the *APOE* SNPs, taking out a single SNP every time. The PRS for each of these lists was calculated and models were fitted as described above. Results from each list were compared and the list with higher beta- and lower p-values compared to the original list was kept. This process was repeated k times (here 16 times), with the best list replacing the original list until neither the beta- nor the p-values could be improved by deleting a single SNP.

2.7. GO category enrichment

SNPs from the A-PRS and oA-PRS lists were analyzed for GO biological process enrichment. If a SNP was located between two genes (i.e., *ZCPW1/NYAP1*), both genes were included in the analysis. GO enrichment analysis was performed using the Enrichr site^{40,41}. Only processes that reached an adjusted p-value of 0.05 were considered significant.

3. Results

3.1. Demographic description of the INSIGHT discovery cohort

Genomic data were obtained from 298 of the original 318 participants included in the INSIGHT pre-AD study. We removed participants with the $APOE\ \epsilon 2/\epsilon 4$ genotype based on the observation that those two alleles show differential effects on amyloid deposition⁴². Genomic data were available from 291 participants. Most participants (208, 71.5%) had amyloid CL values lower than 20, and were therefore classified as amyloid (–). Our population was 61.2% female, and this proportion was similar in amyloid (–) and (+) groups. As expected, participants in the amyloid (+) group were more likely to be $APOE\ \epsilon 4$ carriers (p=0.001) and less likely to be $APOE\ \epsilon 2$ carriers (p=0.002) (Table 2). Additionally, as previously described²⁵, the distribution of CL values did not follow a normal distribution, and there was a weak correlation of these values with age (p=0.036). However, despite this weak correlation, all subsequent models included age as a confounding factor.

3.2. Discovery cohort: PRS association with amyloid status

We used a list of 33 SNPs associated with AD risk⁶ (excluding *APOE*) to generate a first PRS, named A-PRS (Table 1), adjusted for age, sex, and population structure (PC1, PC2, and PC3). We did not observe any significant association between the A-PRS and amyloid status (Figure 1A and Table 3). This lack of association was also observed in an *APOE*-stratified analysis (Figure 1B and Table 3, no interaction between *APOE* status and A-PRS was detected). Therefore, in the discovery cohort, the A-PRS was not associated with amyloid deposition in cognitively unimpaired participants. As expected, due to *APOE* genotype distribution, a model including only *APOE* status showed an association with amyloid status (Table 3) (OR=4.08, [2.17-7.78]).

We then hypothesized that this lack of association may occur because loci associated with AD risk are not linked to amyloid deposition processes. We used an iteration process to select a combination of SNPs leading to a PRS associated with amyloid status. At the end of this process, we obtained an optimized A-PRS (oA-PRS) based on 17 of the original 33 SNPs excluding *APOE* that showed the strongest association with brain amyloid load in the INSIGHT cohort (in grey in Table 1). This association was improved when the model was adjusted for *APOE* status (Figure 1C and Table 3) (OR without APOE= 5.26 [1.71-16.88], OR with APOE= 5.93 [1.85-19.83]). The *APOE*-stratified analysis showed a significant association of the oA-PRS with amyloid status both in ε4 carriers and non-carriers (Figure 1D and Table 3, p-value=0.12 for interaction between oA-PRS and APOE status). A significant correlation between CL values and oA-PRS was observed in the total population (total group: rho=0.13, p=0.03; amyloid (-): rho=0.048, p=0.49; amyloid (+): rho=0.17, p=0.13), which could be caused by the lower CL values in the amyloid (-) population. The large variations observed in the OR among APOE ε4 carriers could be attributed to the small sample size [29 and 32 amyloid (+), and 24 and 23 amyloid (-)

APOE ε4 carriers in INSIGHT and ADNI respectively] compared to the whole population [(83 and 90 amyloid (+), and 208 and 138 amyloid (-) from INSIGHT and ADNI respectively].

To assess differences between A-PRS and oA-PRS, we performed pathway-enrichment analysis (Supplementary eTable 1 for A-PRS and Supplementary eTable 2 for oA-PRS). Biological processes related to $A\beta$ metabolism and oligomerization represented six of the twelve (50%) and five of the seven (71%) significantly enriched pathways when the analysis was performed based on genes assigned to SNPs used in the A-PRS or oA-PRS, respectively (Figure 2).

3.3. Demographic description of the ADNI validation cohort

We selected 230 control subjects from the ADNI cohort⁴³ to validate the oA-PRS. Two subjects with the *APOE* ε 2/ ε 4 genotype were excluded. This ADNI validation cohort had a mean age of 76.6 years and a mean MMSE score of 29.3, similar to the INSIGHT cohort¹⁸. Sex distribution was significantly different between cohorts (p=0.002). Finally, 90 (39.5%) participants from the ADNI cohort were classified as amyloid (+), which was significantly higher than in the INSIGHT cohort (p=0.0002). Additionally, amyloid (–) participants in the validation cohort had lower CL values than amyloid (–) participants in the discovery cohort (p=0.03), whereas the opposite was observed for amyloid (+) participants (p=0.0042). As observed in the discovery cohort, the proportion of ε 4 carriers was higher in the amyloid (+) group (p=0.004) (Table 2). Likewise, the CL values followed a non-normal distribution and no significant correlation was found between age and CL status. However, age was still included in the models to make them comparable with the INSIGHT analyses.

3.4. Validation cohort: PRS association with amyloid status

The two PRSs developed in the discovery cohort were tested in the validation cohort. The A-PRS was not significantly associated with amyloid status in the validation cohort even after stratifying by *APOE* genotype (Figure 3A and Table 3). As expected, *APOE* was also strongly associated with amyloid status (OR= 3.36 [1.73-6.7]).

However, the oA-PRS was significantly associated with amyloid status in the validation cohort (Table 3 and Figure 3C), independent of the addition of *APOE* status in the model. This association remained significant in the *APOE* ε4 carriers, as observed in the INSIGHT discovery cohort (Table 3 and Figure 3D). In this case, however, there was no significant correlation between oA-PRS and CL values (total group: rho=0.046, p=0.49; amyloid (-): rho=-0.075, p=0.36; amyloid (+): rho=-0.0017, p=0.99).

3.5. Demographic description of the EADI cohorts

Finally, we tested the power of the oA-PRS to discriminate between controls and AD patients in the EADI study. After excluding participants for which data for age or APOE genotype were not available and participants that were APOE $\varepsilon 2/\varepsilon 4$, we had a total of 8,515 subjects (Table 2). As expected, the AD group had a higher percentage of APOE $\varepsilon 4$ carriers than the control group (47.9% versus 18.7% respectively).

3.6. EADI cohorts: PRS association with disease status

Two variants from the A-PRS were not present in the TOPMed imputations (*IQCK* rs7185636 and *MAPT* rs2732703) and were thus replaced by proxy variants based on the linkage disequilibrium in the haplotype reference consortium (rs11865116 and rs2532332, respectively).

For the calculation of the A- and the oA-PRSs, the weights used were based on the respective log(OR) from the stage II analyses of the European Alzheimer & Dementia Biobank consortium meta-analysis³³ when available, or otherwise from the stage I analyses (i.e., AC074212.3 rs76320948, ACE rs138190086, IQCK rs11865116, CD33 rs3865444, WWOX/MAF rs62039712). PRS association analyses were adjusted for sex, age, and the first three principal components. We found the A-PRS was associated with disease status (Control or AD) in the EADI cohort whether (OR: 1.68 [1.53-1.85]) or not (OR: 1.66 [1.50-1.83]) we accounted for *APOE* status. Stratified analysis according to *APOE* \$\varepsilon 4\$ genotype showed significant associations independent of the *APOE* group (Table 4). The oA-PRS also was significantly associated with disease status with slightly higher ORs (oA-PRS OR: 2.06 [1.73-2.45], oA-PRS + APOE OR: 1.99 [1.66-2.38]).

4. Discussion

This study identified an optimized PRS associated with amyloid status based on a shortlist of validated AD-risk-associated SNPs (excluding *APOE*) in two independent cohorts of participants without cognitive impairment. Stratified analyses showed that the association prevailed in *APOE* \$\partial 4\$ carriers. This observation indicates that AD-associated genetic risk factors other than *APOE* \$\partial 4\$ may increase the risk of amyloid deposition in *APOE* \$\partial 4\$ carriers who are already at high risk for AD. Interestingly, most of the significant enriched pathways (71.3%) corresponding to the genes assigned to the selected SNPs are linked to APP metabolism and brain amyloid deposition. Finally, we showed that the oA-PRS score restricted to 17 SNPs was also associated with disease status, suggesting its improved utility compared to PRS based on a higher number of SNPs.

Few studies have assessed the association of PRSs with amyloid deposition in AD. Among these studies, only one described a PRS association independent of *APOE* status¹⁵. However, this study included individuals with dementia. Two other studies identified *APOE*-dependent associations of PRS^{9,16}, but only one included cognitively unimpaired participants (with a family history of AD)¹⁶. This heterogeneity in terms of population studied and PRS design makes comparison between studies difficult. A recent retrospective study of a cohort of cognitively unimpaired individuals found that a PRS comprising 39 AD SNPs was associated with an increased likelihood of amyloid positivity in the CSF independent of *APOE* status¹⁷. In addition, this PRS could predict progression to AD dementia¹⁷. This PRS shares 22 loci (16 SNPs) with the A-PRS and 12 loci (8 SNPs) with the oA-PRS. Our study and the recent worm demonstrate that genetic factors beyond *APOE* can impact not only amyloid pathology, but also the risk of developing AD.

While APOE $\varepsilon 4$ carriers have an established higher risk for amyloid deposition and AD, it is of interest to identify risk modifiers, such as the oA-PRS. The oA-PRS is not exclusive for APOE $\varepsilon 4$ carriers, but we found a higher association with amyloid load in APOE $\varepsilon 4$ carriers. On the other hand, the oA-PRS did not correlate with the numerical florbetapir centiloid values in amyloid-positive individuals and APOE $\varepsilon 4$ carriers in the discovery or validation cohorts, and it was thus unable to predict the level of brain amyloid in this sub-group. Additional studies are needed to test this prediction in larger sample sizes. Of note, oA-PRS was correlated to CL values in the total population.

Due to the small number of subjects (9) in the INSIGHT cohort who converted to dementia, we could not evaluate the predictive power of oA-PRS for AD. Therefore, we used the EADI cohort, which includes both AD and control subjects, to evaluate the association of oA-

PRS with disease status. Data on brain amyloid deposition were not available. We found that both PRSs (A-PRS and oA-PRS) were associated with disease status in this population. The association of A-PRS with disease status in the EADI cohort, but not with amyloid deposition in the ADNI and INSIGHT cohorts, suggests that genetic factors in the A-PRS are linked to disease but not to amyloid deposition in elderly asymptomatic subjects. These factors could be potentially linked to the risk of dementia. Nevertheless, the association of oA-PRS to AD suggests that genetic risk factors for brain amyloid deposition could predict disease outcome.

Both the A-PRS and oA-PRS lists were enriched in processes linked to amyloid deposition. However, the A-PRS included pathways that were not directly involved in amyloid deposition, confirming that there are mechanisms linked to AD that may not be associated with amyloid status. Likely, these pathways contribute to later stages of the disease or to processes that occur independently of amyloid deposition in cognitively unimpaired participants. These could include pathways related to neuroinflammation, tau, insulin resistance, oxidative stress, or others.

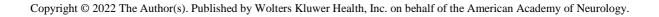
Our study is limited by the sample size of the existing cohorts. Although we acknowledge the value of multiple comparison corrections, here we present the results without correction since the results with correction would not be significant. Nevertheless, we were able to validate the oA-PRS in two independent cohorts with slightly different genetic backgrounds: the INSIGHT cohort composed of white individuals mostly living in Île-de-France, and the ADNI cohort, which is a multiethnic cohort mostly composed of white non-Hispanic Americans. Risk conferred by the £4 variant of *APOE* has been shown to differ across populations, with lower values in populations of African ancestry than in populations of European or Asian ancestries⁴⁴. Additional studies are necessary to validate the oA-PRS in non-white populations. Another

limitation of our study is the age range, as we evaluated people older than 70 who are cognitively unimpaired. In the future, it will be interesting to include younger subjects. This exploratory study will need to be validated in larger cohorts of cognitively unimpaired individuals with brain amyloid imaging.

In conclusion, our findings robustly highlight a PRS excluding *APOE* that is significantly associated with amyloid status in two independent cohorts of cognitively unimpaired individuals. Currently, amyloid load can be measured via plasma or CSF amyloid biomarkers and PET imaging. Genetic risk assessment of amyloid load early in life before any possible detection in plasma or the brain would allow initial screening to establish patient priority for a more detailed follow-up of those at higher risk. Additionally, such assessment would provide stratification for potential preventive or curative treatments based on patient-specific risk factors. A GWAS focusing on cognitively unimpaired participants with significant brain amyloid deposition should unveil new SNPs, some of which could be unrelated to AD, while improving prediction of amyloid load. Beyond genetic data, a combination of omics, genetic, biochemical, and environmental (exposome, diet, microbiome, etc.) features could also allow for a more accurate prediction of amyloid deposition.

Figure legends

Figure 1: PRS in amyloid (+) and amyloid (–) participants from the INSIGHT cohort: A-PRS (A and B) and oA-PRS (C and D). Green-colored violin plots correspond to amyloid (–) participants and orange plots correspond to amyloid (+) participants. Each participant is represented by a colored dot corresponding to their APOE status: dark green for $\varepsilon 2/\varepsilon 2$, orange for $\varepsilon 2/\varepsilon 3$, violet for $\varepsilon 3/\varepsilon 3$, pink for $\varepsilon 3/\varepsilon 4$, and light green for $\varepsilon 4/\varepsilon 4$. For the stratified graphs, subjects who did not carry any $\varepsilon 4$ allele where classified as an "E4 non-carrier", and those who did were classified as an "E4 carrier". A-PRS is not associated with amyloid status in the whole INSIGHT cohort (A) and in the $\varepsilon 4$ carriers (B). The oA-PRS is significantly associated with amyloid status (C) (p=0.005), and this association persists in the $\varepsilon 4$ carriers (p=0.0034) (D); asterisks indicate statistically significant differences (* <0.05, ** <0.001). A-PRS and oA-PRS were not significantly different between APOE statuses among amyloid (+) and (–) participants.



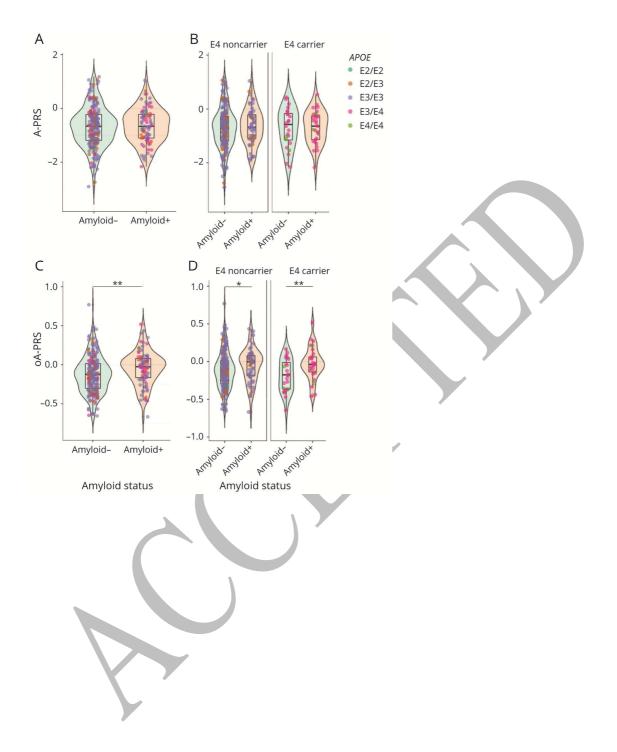


Figure 2: Overlap between enriched GO biological processes in the A-PRS and oA-PRS. In

bold are GO biological processes involved in amyloid pathology.

Regulation of aspartic-type endopeptidase activity involved in amyloid precursor protein catabolic process (GO: 1902959) p = 0.0004

A-PRS

Integrin-mediated signaling pathway (GO: 0007229) p = 0.003

Regulation of neuron death (GO: 1901214) p = 0.0068

Positive regulation of neuron death (GO: 1901216) p = 0.024

Positive regulation of amyloidbeta clearance (GO: 1900223) p = 0.023

Microglial cell activation (GO: 0001774) p = 0.03

Negative regulation of amyloid-beta formation (GO: 1902430)

A-PRS $p = 2.1e^{-6}$ oA-PRS $p = 2.6e^{-4}$

Negative regulation of amyloid precursor protein catabolic process (GO: 1902992)
A-PRS p = 3.8e-8

oA-PRS $p = 2.6e^{-4}$ Regulation of amyloid-beta formation (GO: 1902003)

A-PRS $p = 3.5e^{-7}$ oA-PRS $p = 8.4e^{-4}$

Regulation of cell migration (GO: 0030334) A-PRS p = 0.013 oA-PRS p = 0.009

Regulation of amyloid-beta clearance (GO: 1900221) A-PRS p = 0.0007oA-PRS p = 0.035

Negative regulation of cellular amide metabolic process (GO: 0034249) A-PRS p = 0.007oA-PRS p = 0.034 Negative regulation of protein oligomerization (GO: 0032460)

oA-PRS



Figure 3: PRS in amyloid (+) and amyloid (-) participants from the ADNI cohort: A-PRS (A and B) and oA-PRS (C and D). Green-colored violin plots correspond to amyloid (-) participants while orange plots correspond to amyloid (+) participants. Each participant is represented by a colored dot corresponding to their APOE status: dark green for $\varepsilon 2/\varepsilon 2$, orange for $\varepsilon 2/\varepsilon 3$, violet for $\varepsilon 3/\varepsilon 3$, pink for $\varepsilon 3/\varepsilon 4$, and light green for $\varepsilon 4/\varepsilon 4$. For the stratified graphs, subjects who did not carry any $\varepsilon 4$ allele were classified as "E4 non-carrier", and those who did were classified as "E4 carrier". The A-PRS was not significantly associated with amyloid status in the whole ADNI cohort (A) (p=0.05), or in the APOE stratified groups (B). The oA-PRS is significantly associated with amyloid status in the whole cohort (C) (p=0.049) and in the $\varepsilon 4$ carriers (D) (p=0.012); asterisks indicate statistically significant differences (* <0.05). A-PRS and oA-PRS were not significantly different between APOE statuses among amyloid (+) and (-) participants.

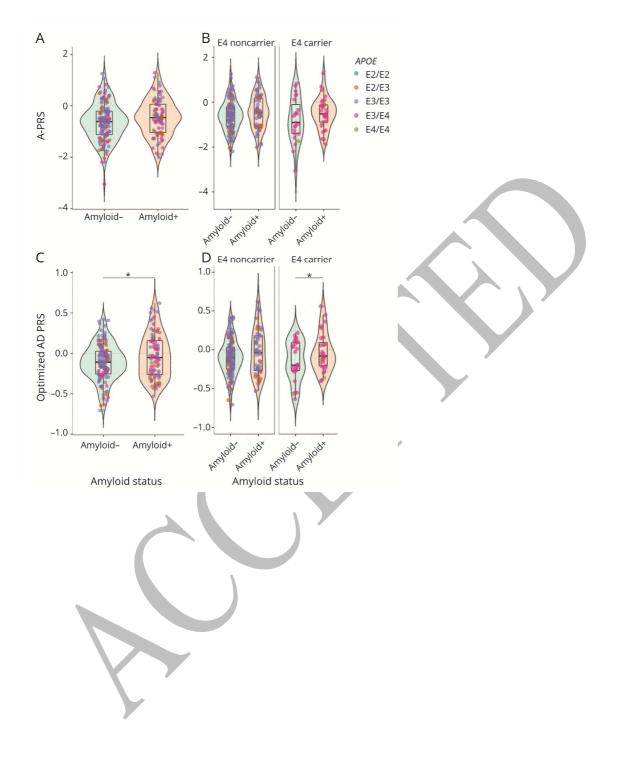


Table 1: List of loci and SNPs for A-PRS and oA-PRS. SNPs were selected from Bellenguez *et al.*⁶ SNPs selected for the optimized A-PRS are highlighted in grey. For each SNP locus, chromosome (Chr), position within the chromosome (hg19), effect allele (EA), other allele (OA) and published odds ratio (OR) are indicated; (1) Substituted by rs9271058 at position 32575406 (2) Substituted by rs12881735 at position 92932828; * No OR has been published for this SNP, and it was excluded in the PRS analysis²⁸.

Table 2: Demographic description of the INSIGHT, ADNI, and EADI cohorts. For each cohort, detailed descriptions of the amyloid (–) and amyloid (+) participants or Controls and AD patients, and total cohorts are reported. For numerical variables, values represent mean, standard deviation and value range. For categorical variables, values include the total number of participants and the percentage in the cohort.

Table 3: Association models fitted for the discovery cohort (INSIGHT) and the validation cohort (ADNI) in ε4 carriers and non-carriers and the unstratified cohort. Models were fitted to binomial models [amyloid (–) and amyloid (+)] using age and sex as confounders and correcting for population structure (with the three first principal components for INSIGHT, first ten principal components for ADNI). Values presented include beta, standard error (SE), p-value, odds ratio (OR) and its 95% confidence interval (CI) for A-PRS (33 SNPs) and oA-PRS (17 SNPs). The *APOE* model includes participants binarized according to ε4 status (ε4 non-carriers, ε4 carriers). Bold values indicate statistically significant comparisons (p<0.05).

Table 4: Association models fitted for the AD cohorts (EADI) in ε4 carriers and non-carriers and the unstratified cohort. Models were fitted to binomial models [Control and AD] using age and sex as confounders and correcting for population structure (with the three first principal components). Values presented include beta, standard error (SE), p-value, odds ratio (OR) and its 95% confidence interval (CI) for A-PRS (33 SNPs) and oA-PRS (17 SNPs). The APOE model includes participants binarized according to ε4 status (ε4 non-carriers, ε4 carriers). Bold values indicate statistically significant comparisons (p<0.05).

Table 1: List of loci and SNPs for A-PRS and oA-PRS. SNPs were selected from Bellenguez et al.⁶ SNPs selected for the optimized A-PRS are highlighted in grey. For each SNP locus, chromosome (Chr), position within the chromosome (hg19), effect allele (EA), other allele (OA) and published odds ratio (OR) are indicated; (1) Substituted by rs9271058 at position 32575406 (2) Substituted by rs12881735 at position 92932828; * No OR has been published for this SNP, and it was excluded in the PRS analysis (30).

Locus	SNP	Chr	Position	EA	OA	OR	Optimized
					V		AD list
ADAMTS4	rs4575098	1	161155392	A	G	1.04	
CRI	rs4844610	1	207802552	A	C	1.17	
BIN1	rs6733839	2	127892810	T	С	1.2	
INPP5D	rs10933431	2	233981912	G	С	0.91	
CLNK	rs6448453	4	11026028	A	G	1.07	
HLA	rs9271192 (1)	6	32578530	С	A	1.1	
OARD1	rs114812713	6	41034000	С	G	1.32	
CD2AP	rs9473117	6	47431284	С	A	1.09	
ZCWPW1/NYAP	rs12539172	7	100091795	T	С	0.92	
1							
EPHA1	rs10808026	7	143099133	A	С	0.9	

PTK2B	rs73223431	8	27219987	T	C	1.1	
CLU	rs9331896	8	27467686	С	T	0.88	
ECHDC3	rs7920721	10	11720308	G	A	1.08	
CELF1/SPI1	rs3740688	11	47380340	G	T	0.92	
MS4A	rs7933202	11	59936926	С	A	0.89	
PICALM	rs3851179	11	85868640	Т	С	0.88	
SORL1	rs11218343	11	121435587	С	T	0.8	
FERMT2	rs17125924	14	53391680	G	A.	1.14	
SLC2A4/RIN3	rs10498633	14	92926952	T	G	0.93	
	(2)						
ADAM10	rs593742	15	59045774	G	A	0.93	
APH1B	rs117618017	15	63569902	T	С	1.1	
IQCK	rs7185636	16	19808163	С	T	0.92	
KAT8	rs59735493	16	31133100	A	G	0.96	
WWOX/MAF	rs62039712	16	79355857	A	G	1.16	
SCIMP/RABEP	rs113260531	17	5138980	A	G	*	
1							
MAPT	rs2732703	17	44353222	G	T	0.73	

ABI3	rs616338	17	47297297	T	С	1.43	
TSPOAP1	rs2632516	17	56409089	С	G	0.94	
ACE	rs138190086	17	61538148	A	G	1.3	
ABCA7	rs3752246	19	1056492	G	C	1.15	
AC074212.3	rs76320948	19	46241841	Т	С	1.18	
CD33	rs3865444	19	51727962	A	С	0.94	
CASS4	rs6024870	20	54997568	A	G	0.88	
ADAMTS1	rs2830500	21	28156856	A	C.	0.93	

2 Table 2: Demographic description of the INSIGHT, ADNI, and EADI cohorts. For each cohort, detailed descriptions of the amyloid

- 3 (-) and amyloid (+) participants or Controls and AD patients, and total cohorts are reported. For numerical variables, values represent
- 4 mean, standard deviation and value range. For categorical variables, values include the total number of participants and the percentage
- 5 in the cohort.

			INSIGHT		ADNI			EADI	
		Amyloid (-)	Amyloid (+)	Total	Amyloid (-)	Amyloid (+)	Total	Controls	AD
		(n=208,	(n=83,	(n=291)	(n=138,	(n=90,	(n=228)	(n=6,215,	(n=2,300,
		71.5%)	28.5%)		60.5%)	39.5%)		73%)	27%)
Age		76.31 ±3.5	77.25 ±3.3	76.4 ±3.5	76.35 ±4.41	77.05 ±4.74	76.62 ±4.55	80.0±7.6	74.3±10.2
		(69.9-86)	(70-86)	(69.9-86)	(69.1-85.7)	(69.2-85.9)	(69.1-85.9)	(40.0-102.3)	(37.0-99.3)
Sex	Fema	130 (62.5%)	48 (57.8 %)	178 (61.2%)	60 (43.5%)	55 (61.1%)	115 (50.4%)	3,749	1,515
	le							(60.3%)	(65.9%)
	Male	78 (37.5%)	35 (42.1%)	113 (38.8%)	78 (56.5%)	35 (38.9%)	113 (49.6%)	2,466	785

								(39.7%)	(34.1%)
Centil	oid	4.56 ±7.84	52.21 ±29.25	18.05 ±27.44	1.65 ±10.83	65.5 ±33.42	26.86 ±38.56		
		(-17.68 —	(20.04—	(-17.68—	(-28.49—	(20.06—	(-28.49—		
		19.64)	139.26)	139.26)	19.91)	202.8)	202.8)		
APO	ε2/ε2	1 (0.48%)	0 (0%)	1 (0.34%)	1 (0.72%)	0 (0%)	1 (0.43%)	38 (0.6%)	7 (0.3%)
E	ε2/ε3	30 (14.4%)	6 (7.2%)	36 (12.4 %)	16 (11.6%)	9 (10%)	25 (10.96%)	772 (12.4%)	127 (5.5%)
	ε3/ε3	153 (73.6%)	48 (57.8%)	201 (69.1%)	98 (71.0%)	49 (54.4%)	147	4,241	1,063
							(64.47%)	(68.2%)	(46.2%)
	ε3/ε4	21 (10.1%)	26 (31%)	47 (16.1%)	22 (15.9%)	30 (33.3%)	52 (22.81%)	1,105	879
								(17.8%)	(38.2%)
	ε4/ε4	3 (1.4%)	3 (3.6%)	6 (2.1%)	1 (0.72%)	2 (2.2%)	3 (1.32%)	59 (0.9%)	224 (9.7%)

Table 3: Association models fitted for the discovery cohort (INSIGHT) and the validation cohort (ADNI) in ε4 carriers and non-carriers and the unstratified cohort. Models were fitted to binomial models [amyloid (–) and amyloid (+)] using age and sex as confounders and correcting for population structure (with the three first principal components for INSIGHT, first ten principal components for ADNI). Values presented include beta, standard error (SE), p-value, odds ratio (OR) and its 95% confidence interval (CI) for the PRS and its p-value for A-PRS (33 SNPs) and oA-PRS (17 SNPs). The APOE model includes INSIGHT participants binarized according to ε4 status (ε4 non-carriers, ε4 carriers). Bold values indicate statistically significant comparisons (p<0.05), italicized values indicate non-significant comparisons (p<0.1).

			APOE A-PRS	A- PRS+APOE	oA-PRS
	ε4 non-carriers	β	0.111		0.314
	amyloid (–	SE	0.159		0.158
):184 amyloid	p-value	0.49		0.047
	(+):54	OR [95% CI]	1.16 [0.76-		3.77 [1.03-14
INSIGH			1.78]		
T	Y	β	0.178		1.187
	ε4 carriers	SE	0.311		0.414
	amyloid (–):24	p-value	0.57		0.004
	amyloid (+):29	OR [95% CI]	1.31 [0.53-		181.6 [7.53
			3.45]		10,674.6]

	Total cohort	β	0.544	0.088	0.110	0.389
		SE	0.126	0.134	0.139	0.136
		p-value	0.000015	0.51	0.43	0.004
		OR [95% CI]	4.08 [2.17-	1.13 [0.79-	1.16 [0.8-	5.26 [1.71-16
		OR [95% CI]	7.78]	1.63]	1.71]	3.20 [1.71 10
	ε4 non-carriers	β		0.103		0.147
	amyloid (–	SE		0.179		0.183
):115 amyloid (+):58	p-value		0.56		0.42
		OR [95% CI]		1.16 [0.69-		1.8 [0.43-7.7
				1.95]		
		β		0.511		0.976
	ε4 carriers	SE		0.34		0.387
ADNI	amyloid (–):23	p-value		0.14		0.012
	amyloid (+):32	OR [95% CI]		1.84 [0.86-		44.94 [3.03-1,
				4.45]		11.51 [5.05 1,
		β	0.520	0.158	0.186	0.304
		SE	0.147	0.149	0.151	0.154
	Total cohort	p-value	0.0004	0.29	0.22	0.049
		OR [95% CI]	2 26 [1 72 6 7]	1.24 [0.83-	1.29 [0.86-	3.38 [1.02-11
		OK [53% CI]	3.36 [1.73-6.7]	1.87]	1.96]	J.J0 [1.U2-11

Table 4: Association models fitted for the AD cohort (EADI) in ε4 carriers and non-carriers and the unstratified cohort. Models were fitted to binomial models [Control and AD] using age and sex as confounders and correcting for population structure (with the three first principal components). Values presented include beta, standard error (SE), p-value, odds ratio (OR) and its 95% confidence interval (CI) for A-PRS (33 SNPs) and oA-PRS (17 SNPs). The APOE model includes participants binarized according to ε4 status (ε4 non-carriers, ε4 carriers). Bold values indicate statistically significant comparisons (p<0.05).

					A DDG
		A DDC	A DDC - ADOE	o A DDC	oA-PRS +
		A-PRS	A-PRS + APOE	oA-PRS	A DOE
					APOE
ε4 non-	β	0.480		0.677	
0111011	,				
carriers	SE	0.061		0.114	
Santuala, 5.05	m1	5.960E-15		4.074E-09	
Controls: 5,05	p-value	5.900E-15		4.0/4E-09	
1	OR [95%	1.62 [1.43-		1.95 [1.56-	
AD: 1,197	CI]	1.82]		2.44]	
	β	0.537		0.669	
ε4 carriers	P	0.557		0.007	
	SE	0.086		0.160	
Controls:					
	p-value	3.475E-10		2.242E-05	
1,164	OD 1050/	1 81 51 45		1 08 51 44	
AD 1,103	OR [95%	1.71 [1.45-		1.97 [1.44-	
AD 1,103	CI]	2.02]		2.69]	
	CIJ	2.02]		2.05]	
Total cohort	β	0.521	0.506	0.721	0.686
Controlo	CE	0.040	0.040	0.000	0.002
Controls:	SE	0.048	0.049	0.089	0.092
6,215	p-value	1.490E-27	1.631E-24	6.004E-16	8.520E-14

AD 2,300	OR [95%	1.68 [1.53-		2.06 [1.73-	
	CI]	1.85]	1.66 [1.50-1.83]	2.45]	1.99 [1.66-2.38]



Appendix 2 – Coinvestigators

Name	Location	Role	Contribution
Hovagim	ICM, Paris	Coinvestigato	INSIGHT-preAD Scientific
Bakardjian,		r=	Committee
PhD			
Habib Benali,	Centre de	Coinvestigato	Major role in the acquisition
PhD	recherche,	r	of MRI data and study
	IUGM,		protocol design
	Montreal		
Hugo Bertin,	GE	Coinvestigato	Major role in the acquisition
MD	Healthcare,	r	of PET data
	Bruxelless		
Joel Bonheur,	IM2A, Paris	Coinvestigato	Major role in the acquisition
MD		r	of Clinical data
Laurie	IM2A, Paris	Coinvestigato	Major role in the acquisition
Boukadida,		r	of Clinical data
MD			
Enrica Cavedo,	Sorbonne	Coinvestigato	Design and conceptualized
PhD	University,	r	study; analyzed the data;
	Paris		Interpreted the data; drafted
			the manuscript for
			intellectual content

Patrizia	Sorbonne	Coinvestigato	Major role in the acquisition
Chiesa, PhD	University,	r	and evaluation of MRI data
	Paris		
Marion Dubois	IM2A, Paris	Coinvestigato	Analyzed the data; revised
	,	\mathbf{r}	the manuscript for
		1	
			intellectual content
Stéphane	ICM, Paris	Coinvestigato	Major role in the acquisition
Epelbaum, MD		r	of Clinical data
PhD			
Geoffroy	IM2A, Paris	Coinvestigato	Major role in the acquisition
Gagliardi		r	of Neuropsychological data
Remy	IM2A, Paris	Coinvestigato	Major role in the
Genthon, MD		r	coordination of the study
Harald	Sorbonne	Coinvestigato	INSIGHT-preAD Scientific
Hampel, MD	University,	r	Committee
	Paris		
Marion Houot	Sorbonne	Coinvestigato	analyzed the data; revised
	University,	r	the manuscript for
	Paris		intellectual content
Aurélie Kas,	APHP, Paris	Coinvestigato	Major role in the acquisition
MD		r	of PET data
Foudil Lamari,	APHP, Paris	Coinvestigato	INSIGHT-preAD Scientific
PhD		r	Committee

Simone Lista,	Sorbonne	Coinvestigato	Interpreted the data; revised
PhD	University,	r	the manuscript for
	Paris		intellectual content
Christiane	ICM, Paris	Coinvestigato	Data Management
Metzinger		r	
Fanny Mochel,	ICM, Paris	Coinvestigato	INSIGHT-preAD Scientific
MD PhD		r	Committee
Francis Nyasse	IM2A, Paris	Coinvestigato	Major role in the
		r	coordination of the study
Catherine	IM2A, Paris	Coinvestigato	Major role in the acquisition
Poisson		r	of data
Marie Revillon	IM2A, Paris	Coinvestigato	Major role in the acquisition
		r	of Neuropsychological data
Antonio Santos	IM2A, Paris	Coinvestigato	Major role in the acquisition
		r	of Clinical data
Katia Santos	IM2A, Paris	Coinvestigato	Major role in the acquisition
Andrade		r	of Clinical data
Marine Sole	IM2A, Paris	Coinvestigato	Major role in the acquisition
	,	r	of EEG data

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WNL-2022-200600_etab2 --http://links.lww.com/WNL/C26

WNL-2022-200600_coinvestigator_appendix3 -- http://links.lww.com/WNL/C27

- 1. Bekris LM, Yu C-E, Bird TD, Tsuang DW. Genetics of Alzheimer disease. *J Geriatr Psychiatry Neurol*. 2010;23(4):213-227. doi:10.1177/0891988710383571
- 2. Gatz M, Reynolds CA, Fratiglioni L, et al. Role of genes and environments for explaining Alzheimer disease. *Archives of general psychiatry*. Feb 2006;63(2):168-74. doi:10.1001/archpsyc.63.2.168
- 3. Strittmatter WJ, Saunders AM, Schmechel D, et al. Apolipoprotein E: high-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc Natl Acad Sci U S A*. Mar 1 1993;90(5):1977-81. doi:10.1073/pnas.90.5.1977
- 4. Sims R, Hill M, Williams J. The multiplex model of the genetics of Alzheimer's disease. *Nat Neurosci*. Mar 2020;23(3):311-322. doi:10.1038/s41593-020-0599-5
- 5. Genin E, Hannequin D, Wallon D, et al. APOE and Alzheimer disease: a major gene with semi-dominant inheritance. *Mol Psychiatry*. Sep 2011;16(9):903-7. doi:10.1038/mp.2011.52
- 6. Bellenguez C, Grenier-Boley B, Lambert JC. Genetics of Alzheimer's disease: where we are, and where we are going. *Curr Opin Neurobiol*. Apr 2020;61:40-48. doi:10.1016/j.conb.2019.11.024

- 7. Adams HH, de Bruijn RF, Hofman A, et al. Genetic risk of neurodegenerative diseases is associated with mild cognitive impairment and conversion to dementia. *Alzheimers Dement*. Nov 2015;11(11):1277-85. doi:10.1016/j.jalz.2014.12.008
- 8. Altmann A, Scelsi MA, Shoai M, et al. A comprehensive analysis of methods for assessing polygenic burden on Alzheimer's disease pathology and risk beyond APOE. *Brain Commun.* 2020;2(1):fcz047. doi:10.1093/braincomms/fcz047
- 9. Chaudhury S, Brookes KJ, Patel T, et al. Alzheimer's disease polygenic risk score as a predictor of conversion from mild-cognitive impairment. *Transl Psychiatry*. May 24 2019;9(1):154. doi:10.1038/s41398-019-0485-7
- 10. Leonenko G, Shoai M, Bellou E, et al. Genetic risk for alzheimer disease is distinct from genetic risk for amyloid deposition. *Ann Neurol*. Sep 2019;86(3):427-435. doi:10.1002/ana.25530
- 11. Rodriguez-Rodriguez E, Sanchez-Juan P, Vazquez-Higuera JL, et al. Genetic risk score predicting accelerated progression from mild cognitive impairment to Alzheimer's disease. *J Neural Transm (Vienna)*. May 2013;120(5):807-12. doi:10.1007/s00702-012-0920-x
- 12. Cruchaga C, Del-Aguila JL, Saef B, et al. Polygenic risk score of sporadic late-onset Alzheimer's disease reveals a shared architecture with the familial and early-onset forms. *Alzheimers Dement*. Feb 2018;14(2):205-214. doi:10.1016/j.jalz.2017.08.013
- 13. Desikan RS, Fan CC, Wang Y, et al. Genetic assessment of age-associated Alzheimer disease risk: Development and validation of a polygenic hazard score. *PLoS Med*. Mar 2017;14(3):e1002258. doi:10.1371/journal.pmed.1002258

- 14. Sleegers K, Bettens K, De Roeck A, et al. A 22-single nucleotide polymorphism Alzheimer's disease risk score correlates with family history, onset age, and cerebrospinal fluid Abeta42. *Alzheimers Dement*. Dec 2015;11(12):1452-1460. doi:10.1016/j.jalz.2015.02.013
- 15. Mormino EC, Sperling RA, Holmes AJ, et al. Polygenic risk of Alzheimer disease is associated with early- and late-life processes. *Neurology*. Aug 2 2016;87(5):481-8. doi:10.1212/WNL.00000000000002922
- 16. Darst BF, Koscik RL, Racine AM, et al. Pathway-Specific Polygenic Risk Scores as Predictors of Amyloid-beta Deposition and Cognitive Function in a Sample at Increased Risk for Alzheimer's Disease. *J Alzheimers Dis.* 2017;55(2):473-484. doi:10.3233/JAD-160195
- 17. Ebenau JL, van der Lee SJ, Hulsman M, et al. Risk of dementia in APOE & carriers is mitigated by a polygenic risk score. *Alzheimer's & dementia (Amsterdam, Netherlands)*. 2021;13(1):e12229. doi:10.1002/dad2.12229
- 18. Dubois B, Epelbaum S, Nyasse F, et al. Cognitive and neuroimaging features and brain beta-amyloidosis in individuals at risk of Alzheimer's disease (INSIGHT-preAD): a longitudinal observational study. *Lancet Neurol*. Apr 2018;17(4):335-346. doi:10.1016/S1474-4422(18)30029-2
- 19. Habert MO, Bertin H, Labit M, et al. Evaluation of amyloid status in a cohort of elderly individuals with memory complaints: validation of the method of quantification and determination of positivity thresholds. *Ann Nucl Med.* Feb 2018;32(2):75-86. doi:10.1007/s12149-017-1221-0
- 20. Xicota L, Ichou F, Lejeune FX, et al. Multi-omics signature of brain amyloid deposition in asymptomatic individuals at-risk for Alzheimer's disease: The INSIGHT-preAD study. *EBioMedicine*. Sep 2019;47:518-528. doi:10.1016/j.ebiom.2019.08.051

- 21. Klunk WE, Koeppe RA, Price JC, et al. The Centiloid Project: standardizing quantitative amyloid plaque estimation by PET. *Alzheimers Dement*. Jan 2015;11(1):1-15.e1-4. doi:10.1016/j.jalz.2014.07.003
- 22. platform C.
- 23. Navitsky M, Joshi AD, Kennedy I, et al. Standardization of amyloid quantitation with florbetapir standardized uptake value ratios to the Centiloid scale. *Alzheimers Dement*. Dec 2018;14(12):1565-1571. doi:10.1016/j.jalz.2018.06.1353
- 24. Amadoru S, Doré V, McLean CA, et al. Comparison of amyloid PET measured in Centiloid units with neuropathological findings in Alzheimer's disease. *Alzheimer's Research & Therapy*. 2020/03/04 2020;12(1):22. doi:10.1186/s13195-020-00587-5
- 25. La Joie R, Ayakta N, Seeley WW, et al. Multisite study of the relationships between antemortem [(11)C]PIB-PET Centiloid values and postmortem measures of Alzheimer's disease neuropathology. *Alzheimers Dement*. Feb 2019;15(2):205-216. doi:10.1016/j.jalz.2018.09.001 26. database A.
- 27. Vascular factors and risk of dementia: design of the Three-City Study and baseline characteristics of the study population. *Neuroepidemiology*. Nov-Dec 2003;22(6):316-25. doi:10.1159/000072920
- 28. Lambert JC, Heath S, Even G, et al. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat Genet*. Oct 2009;41(10):1094-9. doi:10.1038/ng.439
- 29. Blauwendraat C, Faghri F, Pihlstrom L, et al. NeuroChip, an updated version of the NeuroX genotyping platform to rapidly screen for variants associated with neurological diseases. *Neurobiol Aging*. Sep 2017;57:247 e9-247 e13. doi:10.1016/j.neurobiolaging.2017.05.009

- 30. Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: rising to the challenge of larger and richer datasets. *Gigascience*. 2015;4:7. doi:10.1186/s13742-015-0047-8
- 31. McCarthy S, Das S, Kretzschmar W, et al. A reference panel of 64,976 haplotypes for genotype imputation. *Nat Genet*. Oct 2016;48(10):1279-83. doi:10.1038/ng.3643
- 32. Kunkle BW, Grenier-Boley B, Sims R, et al. Genetic meta-analysis of diagnosed Alzheimer's disease identifies new risk loci and implicates Abeta, tau, immunity and lipid processing. *Nat Genet*. Mar 2019;51(3):414-430. doi:10.1038/s41588-019-0358-2
- 33. Bellenguez C, Küçükali F, Jansen I, et al. New insights on the genetic etiology of Alzheimer's and related dementia. *medRxiv*. 2020:2020.10.01.20200659. doi:10.1101/2020.10.01.20200659
- 34. Karczewski KJ, Francioli LC, Tiao G, et al. The mutational constraint spectrum quantified from variation in 141,456 humans. *Nature*. 2020/05/01 2020;581(7809):434-443. doi:10.1038/s41586-020-2308-7
- 35. Taliun D, Harris DN, Kessler MD, et al. Sequencing of 53,831 diverse genomes from the NHLBI TOPMed Program. *Nature*. 2021/02/01 2021;590(7845):290-299. doi:10.1038/s41586-021-03205-y
- 36. Das S, Forer L, Schönherr S, et al. Next-generation genotype imputation service and methods. *Nat Genet*. Oct 2016;48(10):1284-1287. doi:10.1038/ng.3656
- 37. Jansen E, Savage JE, Watanabe K, et al. Genome-wide meta-analysis identifies new loci and functional pathways influencing Alzheimer's disease risk. *Nat Genet*. Mar 2019;51(3):404-413. doi:10.1038/s41588-018-0311-9

- 38. Team RC. R: A language and environment for statistical computing. *R Foundation for Statistical Computing, Vienna, Austria.* 2020;
- 39. Lambert JC, Ibrahim-Verbaas CA, Harold D, et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nature genetics*. 2013;45(12):1452-1458. doi:10.1038/ng.2802
- 40. Chen EY, Tan CM, Kou Y, et al. Enrichr: interactive and collaborative HTML5 gene list enrichment analysis tool. *BMC bioinformatics*. Apr 15 2013;14:128. doi:10.1186/1471-2105-14-128
- 41. Kuleshov MV, Jones MR, Rouillard AD, et al. Enrichr: a comprehensive gene set enrichment analysis web server 2016 update. *Nucleic acids research*. Jul 8 2016;44(W1):W90-7. doi:10.1093/nar/gkw377
- 42. Grothe MJ, Villeneuve S, Dyrba M, Bartrés-Faz D, Wirth M. Multimodal characterization of older APOE2 carriers reveals selective reduction of amyloid load. *Neurology*. Feb 7 2017;88(6):569-576. doi:10.1212/wnl.0000000000003585
- 43. Weiner MW, Veitch DP, Aisen PS, et al. 2014 Update of the Alzheimer's Disease Neuroimaging Initiative: A review of papers published since its inception. *Alzheimers Dement*. Jun 2015;11(6):e1-120. doi:10.1016/j.jalz.2014.11.001
- 44. Rajabli F, Feliciano BE, Celis K, et al. Ancestral origin of ApoE ε4 Alzheimer disease risk in Puerto Rican and African American populations. *PLoS genetics*. Dec 2018;14(12):e1007791. doi:10.1371/journal.pgen.1007791



Association of APOE-Independent Alzheimer Disease Polygenic Risk Score With Brain Amyloid Deposition in Asymptomatic Older Adults

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